EFFECTS OF L-ARGININE SUPPLEMENTATION ON INDOMETHACIN-INDUCED GASTRIC ULCERATION IN RATS

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ABSTRACT
Over the last decade emphasis on therapeutic measures has shifted from curative medicine to preventive medicine. In the domain of ulcer and its treatment, not much can be said to have been achieved in terms of preventing ulcer. Most therapies to date have focused on isolating and treating the aggressive factor(s) responsible for the ulcer. A major find and current trend in ulcer management now is the role of nitric oxide (NO) and its agonists such as L-Arginine in ameliorating gastric ulcers. The current study focuses on the possible effects of L-Arginine supplementation on indomethacin-induced gastric ulcerations in rats.

Male Wistar albino rats weighing between 150-180g were used for the study. Animals were divided according to the following experimental groups: Group I (Control) – saline, 8ml/kg, Group II- L-Arginine 1g/kg B.W for 3 days, Group III- L-Arginine 1g/kg B.W for 7 days. Studies carried out included the effect of L-arginine supplementation on indomethacin-induced gastric ulcerations; determination of ulcer dimensions (area, depth and width) and histological profile of the gastric mucosa.

The result showed that arginine supplementation possesses capabilities of reducing ulcer formation in indomethacin-induced rats. Arginine supplementation has an ulcer reducing effect as evident by the lower ulcer scores of the supplemented groups when compared with the Control. The 3-day supplemented group had a significantly lower ulcer score (p<0.05). Ulcer area and width were significantly reduced in the 3-day supplemented group (p<0.05) while the ulcer depth was significantly reduced in both supplemented groups (p<0.01). Histological examination of the gastric tissues showed that the 3-day supplemented group had greater reduction in the level of gastric disturbance than both Control group and the 7-day supplemented group. We proposed that a rapid angiogenetic activity in response to arginine supplementation which then wanes over time may be responsible for this pattern. This study provides evidence that arginine supplementation is beneficial in reducing the risk of gastric ulcer induced by NSAIDS and this is possibly mediated by direct enhancement of angiogenetic activity around the disturbed gastrum.

Key words: Arginine, gastric ulcer, nitric oxide, cGMP, ulcer
INTRODUCTION
One of the major paradigm shifts in the field of medicine is the refocusing of medical therapies from curative medicine to preventive medicine. In the domain of ulcer and its treatment, not much can be said to have been achieved in terms of preventing ulcer; no thanks to the multiple causative sources (aggressive factors) which has made prevention a herculean task. Most therapies to date have focused on isolating and treating the aggressive factor(s) responsible for the ulcer while others have generally relied on careful and strict abstinence from harmful causative sources. A major find and current trend in ulcer management is the role of nitric oxide (NO) and its precursor in ameliorating gastric ulcers (Elliot et al, 1995; Medeiros et al, 2008). Several studies have documented the gastro-protective effects of NO against non-steroidal anti-inflammatory drugs (NSAID)-induced gastric ulceration. By extension L-Arginine, a precursor of NO has also been implicated in having positive effects on gastric ulceration. Such positive benefits have included increased prostaglandin secretion, reduction in the amount of acid secreted and enhanced anti-oxidant action. Earlier studies have shown that pre-treatment with a single dose of NO synthase or its precursor ameliorates NSAID-induced gastropathy in rats (Ajiboye and Oluwole, 2012). There is however a paucity of data on the long-term benefits, if any, that may be derived from regular or supplementary use of NO precursors. This is why the current study focuses on the possible effects of L-Arginine supplementation on indomethacin-induced gastric ulcerations in rats.

MATERIALS AND METHODS
ANIMALS- Male Wistar albino rats weighing between 150-180g were used for the study. They were procured from the Babcock University animal house, Babcock University, Nigeria. The animals were fed with standard rat and mice pellets (Ladokun feeds, Ibadan) and given free access to water. Animals were divided according to the following experimental groups: Group I (Control) – saline, 8ml/kg, Group II- L-Arginine 1g/kg B.W for 3 days, Group III- L-Arginine 1g/kg B.W for 7 days
Studies carried out are:
Study 1 – Effect of L-arginine supplementation on indomethacin-induced gastric ulcerations.
Study 2 – Determination of ulcer dimensions (area, depth and width)
Study 3 - Histological profile of gastric mucosa

CHEMICALS- Indomethacin and L-arginine were purchased from Sigma Aldrich chemicals, Germany.

EXPERIMENTAL PROCEDURES
Ulcer Induction
Indomethacin-induced ulceration model was adapted for this study (Wallace et al, 2000). Animals were fasted for 24 hours prior to experimentation but had free access to water. The Control group received Indomethacin (40mg/kg bw, p.o). Group II and Group III animals also received indomethacin (40mg/kg bw, p.o) at the end of their supplementation periods respectively. All animals were sacrificed 4hours after indomethacin administration with ether overdose; the stomachs were removed and assessed for ulcer lesions (Elegbe, 1978).

Determination of Ulcer scores
This is based on a modified method of Alphin and Ward (1967). Ulcers were independently assessed and scored by two observers using the following criteria- Normal stomach (0),
Punctuate/pin-point ulcer (0.5), two or more small haemorrhagic ulcers (1), ulcers greater than 2mm (2).

**Index of Ulceration**

Paul’s Index was used as ulcer index (Martin-Aragon et al., 1994). This index is expressed as the formula:

\[ M \times N/100 \]

where \( M \) = mean number of ulcers per rat in the group, \( N \) = percentage of rats with ulcer in the group.

**Percentage Inhibition**

\[ \% \text{ Inhibition} = \left( \frac{I_C - I_t}{I_C} \right) \times 100 \]

where \( I_C \) is the ulcer index for Control group, \( I_t \) is the ulcer index for the treated group.

**Determination of Ulcer Dimensions (Area, Depth and Width)**

Ulcer dimension (Area, Depth and Width) determinations were determined using micrometric software, Motic 2000. The stomach samples were flattened and carefully sandwiched between two layers of transparent plastic folders and then scanned with the Motic 2000 image software.

**Histological Assessment**

Histological profiling of the ulcerated gastric mucosa was based on the method described by Sheehan and Hrapchak (1987) using Periodic Acid Schiff staining (PAS). Stomachs fixed with 10% formalin and embedded in paraffin were sectioned at 5µm in an automated microtome. The gastric tissue integrity (mucosa-submucosa) was assessed for damage.

**STATISTICAL ANALYSIS**

Data were expressed as Mean ± SEM. Statistical difference between test groups and control group was calculated using the student’s t-test. \( p<0.05 \) was considered as significant.

**RESULTS AND DISCUSSION**

**TABLE 1:** Effect of L-arginine supplementation on ulcer indices

<table>
<thead>
<tr>
<th>Treatment Group</th>
<th>Mean Ulcer score</th>
<th>Paul’s Index</th>
<th>Percentage Inhibition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group I - Control</td>
<td>13.7 ± 1.47</td>
<td>13.7</td>
<td>-</td>
</tr>
<tr>
<td>(Saline 8ml/kg)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Group II - 1g/kg Arginine (3days)</td>
<td>8.12 ± 2.82</td>
<td>4.06</td>
<td>70.4</td>
</tr>
<tr>
<td>Group III - 1g/kg Arginine (7days)</td>
<td>11.4 ± 3.69</td>
<td>8.55</td>
<td>37.6</td>
</tr>
</tbody>
</table>

Data expressed as Mean±SEM; \( n=8 \). *- significant at \( p<0.05 \) when compared with Control; **- significant at \( p<0.01 \) when compared with Control. NB- 50% of Group II and 25% of Group III failed to develop ulcer.

**TABLE 2:** Ulcer Dimensions

<table>
<thead>
<tr>
<th>Treatment Group</th>
<th>Area (µm²)</th>
<th>Width (µm)</th>
<th>Depth (µm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group I - Control</td>
<td>98.24 ± 6.61</td>
<td>78.22 ± 5.31</td>
<td>2687 ± 250</td>
</tr>
<tr>
<td>(Saline 8ml/kg)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Group II - 1g/kg Arginine (3days)</td>
<td>40.10 ± 2.33**</td>
<td>54.81 ± 3.10*</td>
<td>20.88 ± 2.41**</td>
</tr>
</tbody>
</table>
### Table 1

<table>
<thead>
<tr>
<th>Group</th>
<th>III-1g/kg</th>
<th>104 ± 1.21</th>
<th>66.16 ± 1.66</th>
<th>45.60 ± 3.62**</th>
</tr>
</thead>
</table>

Data expressed as Mean±SEM; n= 8. *- significant at p<0.05 when compared with Control; **- significant at p<0.01 when compared with Control.
FIG. 1: Control animal histology.

Mucosa and submucosa exfoliation in Control animal (arrowed). Gastric erosion extends into the muscularis externa (x40). White markings are labels for ulcer depth measurement.

FIG. 2A: Group II animal (3-day supplementation) histology I

Normal gastric gland and mucosa structure in Group II animals (x40). White markings are labels for ulcer depth measurement.
FIG 2B: Group II animal (3-day supplementation) histology II

Mucosa and submucosa exfoliation in (arrowed) (x40).

FIG 3A: Group III animal (7-day supplementation) histology I

Slight apical glandular erosions (arrowed) with a fairly intact sub-mucosa (x40). White markings are labels for ulcer depth measurement.
The quest to conquer gastric ulcer has led to the discovery and administration of various kinds of therapies over the years. It is an accepted fact that improved vascular circulation enhances the gastrum’s defensive prospects against ulcer (Hirose et al, 1991). Nitric oxide, an endogenous substance has been known to improve microcirculation around ulcers. Furthermore, the NO/cGMP pathway has been implicated in the pathogenesis of ulcers by several authors (Duffin et al., 2008; Santos et al., 2005). Recent evidence has also shown that inducible NO promotes angiogenesis by enhancing vascular endothelial growth factor (VEGF) synthesis (Zandifar et al, 2012). L-arginine is an NO precursor with similar capabilities as NO. It was therefore proposed that arginine supplementation may be beneficial in preventing gastric ulcer formation.

The result of this study showed that arginine supplementation possesses capabilities of reducing ulcer formation in indomethacin-induced rats. Table 1 shows that arginine supplementation has an ulcer reducing effect as evident by the lower ulcer scores of the supplemented groups when compared with the Control. The 3-day supplemented group had a significantly lower ulcer score (p<0.05). This ulcer reducing effect is better seen in the ulcer inhibition index in which the 2 supplemented groups had a highly significant inhibition index when compared with the Control group (p<0.01). Only 50% of the 3-day supplemented group and 75% of the 7-day supplemented group developed ulcer. These figures are in line with studies that have shown that NO-donors ameliorate gastric mucosa damage caused by NSAID agents (Nakamura 2003, Khattab et al., 2001).

Table 2 shows the effect of arginine supplementation on ulcer dimensions. Ulcer area and width were significantly reduced in the 3-day supplemented group (p<0.05) while the ulcer depth was significantly reduced in both supplemented groups (p<0.01). As earlier mentioned, it
may be possible that the level of angiogenesis fostered by L-arginine may be responsible for the reduction in ulcer depth.

Oral administration of 40mg/kg indomethacin to rats provoked deep and severe disturbance of the gastric mucosa. The standard damage pattern usually is epithelial exfoliations that most times extend down into the muscularis mucosae. Histological examination of the gastric tissues however showed a reduction in the level of disturbance especially in the 3-day supplemented group (Fig. 2A, 2B) when compared with the Control group (Fig. 1). While it may be expected that protection against gastric ulcer formation will increase with increasing days of supplementation, this however was not the observed case. The 7-day supplemented group presented with more severe mucosa damage (Fig. 3A, 3B) when compared with the 3-day group. With a similar pattern observed in the ulcer inhibition index and the ulcer depth profile, we postulate that a rapid angiogenetic activity in response to arginine supplementation which then wanes over time is responsible for this pattern. This rapid angiogenesis may also be a major factor in the non-development of ulcer in 50% of the animals in the 3-day supplemented group and as the effect wanes more of the animals developed ulcer, as observed in the 7-day supplemented group. Further studies are planned on longer supplementation days with varying doses of arginine while also incorporating direct angiogenetic studies.

CONCLUSION
This study provides evidence that arginine supplementation is beneficial in reducing the risk of gastric ulcer induced by NSAIDS and this is possibly mediated by direct enhancement of angiogenetic activity around the disturbed gastrum.

REFERENCES


